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# Buffers

### PTx.2% (1L)

* 100mL PBS 10X (900mL H2O)
* 2mL Tween20 (iDISCO uses tritonX)

### PTwH (1L)

* 100mL PBS 10X
* 2mL Tween-20
* 1mL of 10mg/mL Heparin stock solution

### Blocking/Permeabilizing Solution (50mL)

* 42mL PTx.2%
* 3mL of Donkey Serum
* 5mL of DMSO
* 0.1% Sodium azide (0.05 g)
* raise to 1% Tween20 (add 0.4mL)

## Secondary antibodies

Do not use chemical dyes, ALEXA dyes or fluorescent proteins only

# Sample Collection

1. Anesthetize the mouse.

2. Perfuse with 10mL PBS.

3. Perfuse with 10mL 4%PFA/PBS.

4. Dissect the brain/organ and trim to the appropriate size.

5. Fix in 1xPBS/4%PFA at 4°C, 48hr with rocking.

6. Wash in PBS with shaking: RT 30min x 3times.

Add screen shots for clarity:

# Immunolabeling

After fixation and wash:

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| Tissue | Bleach 5% H2O2 in PBS1 | Block/Perm soln | 1° antibody, in Block soln w/o Gt serum | 2nd antibody, in Block soln w/o Gt serum |
| Whole Brain (Klingberg) | O/N rock, wash PTx.2% | 7 days rock @ RT | 7 days rock, wash PTwH @ RT 5x 1 hr each | 1 day rock, wash PTwH @ RT 5x 1 hr each |
| Whole Brain (iDISCO+) |  | 2 days |  | 2 days |

1. Bleach in fresh 3%H2O2 in PBS (1 volume 30% H2O2 to 5 volumes PBS), overnight at RT°C.

# Clearing tissue

After immunolableing:

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| Tissue | 50% EtOH + 2% Tween (pH 9) | 75% EtOH + 2% Tween (pH 9) | 2x 100% EtOH + 2% Tween (pH 9) | 2x ECi |
| Brain | 24hr | 24hr | 24hr each | 4hr each |
| Whole Brain (iDISCO+) |  |  |  |  |

# Tissue check

How clear does it look? Yellowish?

# References

* Hyperlink or paper citation
* <https://idiscodotinfo.files.wordpress.com/2015/04/whole-mount-staining-bench-protocol-methanol-dec-2016.pdf>
* Klingberg et al., Fully Automated Evaluation of Total Glomerular Number and Capillary Tuft Size in Nephritic Kidneys Using Lightsheet Microscopy, 2017.